

ENZYMATIC REGIOSPECIFIC OXIDATIVE CONVERSION OF 5-THIO TO 5-OXO BY
 BAKER'S YEAST IN 5,11-DITHIOPYRROLO[1,4]BENZODIAZEPINE ANTIBIOTICS

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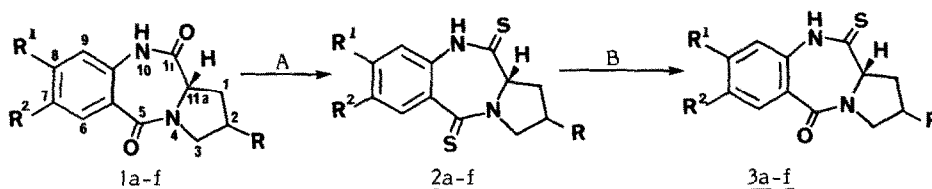
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Abstract Bio-oxidative conversion of 5-thio to 5-oxo by baker's yeast in 5,11-dithio-pyrrolo[2,1-c][1,4]benzodiazepines with regiospecificity is described.

It is well known that baker's yeast (*Saccharomyces cerevisiae*) reduces the carbonyl compounds regio- and stereoselectively.¹ It also reduces the aromatic nitro compounds to anilines,² while the reduction or oxidation of other functional groups has not been much attended. In continuation of our studies on the application of enzymes as biocatalysts in organic synthesis,³ we herein report a regiospecific bio-oxidative conversion of (11aS)-2-acetoxy-1,2,3,10,11,11a-hexahydro-5H-pyrrolo[2,1-c][1,4]benzodiazepine (PBD) 5,11-dithiones (**2**) by baker's yeast to PBD 5-one-11-thiones (**3**) as useful synthons in the synthesis of DNA binding PBD antibiotics.⁴

The precursors PBD-5,11-dithiones⁵ (**2**) were obtained by the reaction of the corresponding dilactams⁶ (**1**) with Lawesson's reagent. Compounds (**2**) upon incubation with baker's yeast gave PBD 5-one-11-thiones (**3**). This novel method of biooxidative conversion exhibited generality with various substituted PBD's having 2 α or 2 β acetoxy groups in 72 to 87% yields.



R = OCOMe

a, R¹ = R² = H, 2 α ; b, R¹ = R² = H, 2 β ; c, R¹ = OH, R² = OMe, 2 α ;
 d, R¹ = OH, OMe, 2 β ; e, R¹ = OCH₂Ph, R² = OMe, 2 α ; f, R¹ = OCH₂Ph, OMe, 2 β

A. Lawesson's Reagent, toluene, 110°C, 3 h (83-88%)

B. Baker's yeast, EtOH, buffer (pH 11.2), 37°C, 2 days

In a typical reaction: to **2a** (150 mg) dissolved in ethanol (10 ml) and 0.1 M phosphate buffer (50 ml; pH 11.2) was added baker's yeast (Sigma, Type I; 1.5 g). Incubation was carried out under aerobic condition at 37°C for 2 days with gentle shaking. The incubation mixture was extracted twice with ethyl acetate (100 ml). The extract was dried over Na₂SO₄ and evaporated to dryness under reduced pressure. The

residue was subjected to column chromatography, chloroform-methanol (96:4) to give PBD 5-one-11-thione (3a)⁷ in 78% yield: m.p. 96-98°C. A control incubation using a boiled yeast preparation afforded 98% recovery of the starting compound.

During our attempts for the non-enzymatic hydrolytic desulfurization⁸ of PBD 5,11-dithiones (2) the desired monothio compounds (3) or even PBD 5,11-diones (1) were not found. Whereas, in these reactions the decomposed material was isolated.

It has been observed in our earlier studies⁵ that in the preparation of PBD 5-one-11-thione (a key intermediate in one of the routes for the synthesis of carbinol-amine PBD antibiotics) by well established procedures^{4c} usually produced 20-25% of the 5,11-dithio compounds. As such this enzymatic procedure is also of value in the selective bio-oxidative conversion of these dithio side-products to (3).

Hence, the present study demonstrates a mild and convenient method for the regio-specific bio-oxidative conversion of PBD 5,11-dithiones to PBD 5-one-11-thiones employing baker's yeast for the first time and in general has a wide potential utility in the selective bio-oxidative conversion of positionally different dithio to monothio compounds.

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- Analytical and spectroscopic data were satisfactory. Selected data for 2a: m.p. 122-124°C; IR (KBr) 3420, 1775, 1590 cm^{-1} ; ^1H NMR (300 MHz, $\text{CDCl}_3 + d_6\text{-DMSO}$), δ 2.09 (s, 3H), 2.57 (ddd, 1H, $J=14.0, 9.3, 9.4$ Hz), 3.75 (br d, 1H, $J=14.3$ Hz with fine coupling), 4.15 (br d, 1H, $J=4.4$ Hz), 4.36 (dd, 1H, $J=9.3, 5.5$ Hz), 4.45 (br d, 1H, $J=8.3$ Hz), 5.35 (m, 1H), 7.19-7.47 (m, 3H), 8.28 (d, 1H, $J=8.1$ Hz), 11.3 (br s, 1H), ^{13}C NMR 21.6, 35.0, 60.1, 64.7, 70.9, 121.4, 121.5, 125.6, 125.9, 132.1, 133.3, 133.9, 134.6, 170.7, 192.6, 199.8; MS 306 (M^+ , 100), 3a: IR (KBr) 3415, 1770, 1740, 1610 cm^{-1} ; ^1H NMR (300 MHz, $\text{CDCl}_3 + d_6\text{-DMSO}$), δ 2.06 (s, 3H), 2.49 (ddd, 1H, $J=14.6, 9.4, 5.5$ Hz), 3.65 (br d, 1H, $J=13.2$ Hz with fine coupling), 3.84 (br d, 1H, $J=13.2$ Hz), 4.01 (dd, 1H, $J=8.1, 5.5$ Hz), 4.29 (d, 1H, $J=9.4$ Hz), 5.30 (m, 1H), 7.12 (d, 1H, $J=8.1$ Hz), 7.32-7.57 (m, 2H), 8.07 (d, 1H, $J=8.1$ Hz), 10.15 (br s, 1H), ^{13}C NMR 21.5, 35.0, 52.8, 60.0, 71.3, 120.9, 125.4, 126.6, 131.6, 133.0, 133.9, 136.0, 165.8, 170.8, 202.4; MS 290 (M^+ , 51).
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